EIV22024E

ExCell Bio

OptiVitro® 293 Serum-free Medium TransExp HE02

For Research and Manufacturing Use

Not Intended for Diagnostic and Therapeutic Use

User Manual

Catalog Number	HE000-N051
Catalog Number	HE000-N052
Catalog Number	HE000-N061
Catalog Number	HE000-N062
Catalog Number	HE000-N063
Catalog Number	HE000-N064



PRODUCT DESCRIPTION

OptiVitro® 293 Serum-free Medium TransExp HE02 is a chemically-defined medium without any animal-derived components or protein, specifically designed for protein production. It is suitable for expanding HEK293-derived suspension cell lines, such as 293T and 293F cells. This medium allows for efficient growth and transfection of multiple suspension 293 cell lines, with a significant increase in protein production when combined with OptiVitro® 293 Serum-free Feed Medium HA02.

SPECIFICATION, STORAGE AND TRANSPORTATION REQUIREMENT

Name	Cat.#	Specification	Storage	Transportation	Shelf Life
OptiVitro® 293 Serum-free Medium TransExp HE02	HE000-N051	500 mL	Store at 2-8°C. Protect From Light.	<25°C，Protect From Light.	12 months
	HE000-N052	1000 mL	Store at 2-8°C. Protect From Light.	<25°C，Protect From Light.	12 months
OptiVitro® 293 Serum-free Medium TransExp HE02(Powder)	HE000-N061	1 L	Store at 2-8°C. Dark and dry.	<10°C，Protect From Light	24 months
	HE000-N062	10 L	Store at 2-8°C. Dark and dry.	<10°C，Protect From Light	24 months
	HE000-N063	100 L	Store at 2-8°C. Dark and dry.	<10°C，Protect From Light	24 months
	HE000-N064	500 L	Store at 2-8°C. Dark and dry.	<10°C，Protect From Light	24 months

PERFORMANCE, APPLICATION AND HANDLING RECOMMENDATIONS

1. Store cell culture medium in a dark environment, ideally in colored packaging, to protect it from light exposure.

2. Avoid prolonged exposure to fluorescent or other types of lighting during transport to prevent discoloration.

3. Implement thorough cleaning and sterilization methods for transport to sterile areas; avoid UV sterilization.

4. Switch off UV lamps when transferring through UV-sterilized windows.

INSTRUCTION FOR USE

Cell Culture

1. Incubate cells at 37℃ in a humidified atmosphere with 5-8% CO_2, using an orbital shaker platform rotating at either 125 rpm (with a 19 mm orbital diameter) or 95 rpm (with a 50 mm orbital diameter).

2. To culture 293 cells in a shake flask, seed a suspension of cells at a concentration of 0.3-1×10^6 cells/mL, with a recommended culture volume of 20-30 mL in a 125 mL shake flask. Subculture cells every 48-72 hours or when cell density reaches 4.0-6.0×10^6 cells/mL.

3. If cells have just been recovered from a frozen state or cultured in other brands of medium, subculture them for three passages before use.

Medium preparation

To prepare 1L of liquid medium from OptiVitro® 293 Serum-free Medium TransExp HE02 (Powder) (ExCell Bio, HE000-N06#), follow these steps:

1. Start with a clean vessel and add approximately 800 mL of water.

2. Slowly add 21.94 g of medium powder while stirring continuously. Mix for about 20 minutes.

3. Adjust the pH with 5 mol/L NaOH to 8.5 ~ 8.8 (about 4.5-5.0 mL). Mix for 10 minutes.

4. Slowly add 2.2 g of sodium bicarbonate powder and mix for 10 minutes.

5. Adjust the pH to 6.9-7.2 with 6 mol/L HCl solution. Mix for another 10 minutes.

6. Add water to reach a final volume of 1L, and continue stirring for an additional 5 minutes.

7. Sterilize by 0.22 μm PES membrane filtration.

8. Store the medium in a cool, dark place at 2℃ to 8℃ for up to 12 months.

Recommendation of Transfection

1. Following cell recovery, subculture cells consistently at least three times to ensure cell viability exceeds 90%.

2. The day prior to transfection, seed cells at an inoculation density of 1.7×10^6 cells/mL in fresh medium.

3. On the day of transfection, adjust the cell volume to 18 mL with fresh medium, maintaining a final transfection density of around 3.3×10^6 cells/mL.

4. **Prepare PEI/DNA complex:** This protocol details the transfection process with a culture volume of 20 mL, a cell density of 3 x 10^6 cells/mL, a DNA concentration of 1.5 μg/mL, and a DNA:PEI ratio of 1.5:4.

1）Dilute 80 μg of PEI Max with 1 mL of OptiVitro® 293 Serum-free Medium TransExp HE02, incubate the mixture at room temperature for 5 minutes.

2）Dilute 30 μg of DNA with 1 mL of OptiVitro® 293 Serum-free Medium TransExp HE02, incubate the mixture at room temperature for 5 minutes.

3）Add the PEI Max solution to the DNA solution to create the PEI/DNA complex, thoroughly mix the solutions and incubate at room temperature for an additional 10 minutes to allow the complex to form.

5. Gently introduce 2 mL of the PEI/DNA complex into the cell suspension, ensuring thorough mixing.

6. After 18-24 hours post-transfection, add 5% volume of OptiVitro® 293 Serum-free Feed Medium HA02, supplemented with 6g/L glucose.

7. Harvest the culture supernatant on day 5 post-transfection, or on day 7 if continued with additional 3g/L glucose supplementation.

If larger volumes of cell transfection are needed, the recommended amount of the reagents are listed below:

Table 1. Recommended dosage for various transfection specifications

Cell culture vessel	125 mL	500 mL	1 L	Remark
Amount of cell ($\times 10^6$ cells)	120	600	1200	cell density 6×10^6 cells/mL
OptiVitro® 293 Serum-free Medium TransExp HE02 (mL)	18	90	180	Initial culture volume
DNA diluent (mL)	1	5	10	
PEI diluent (mL)	1	5	10	
DNA (μg)	30	150	300	DNA: PEI=1.5:4
PEI Max (μg)	80	400	800	
OptiVitro® 293 Serum-free Feed Medium HA03 (mL)	1	5	10	5% of the initial transfection volume
Final culture system (mL)	~21	~105	~210	/

Table 2. Related products

Product Name	Cat.#	Specification
OptiVitro® Glucose Solution	M101381C	10 mL Liquid
	M101382C	100 mL Liquid
OptiVitro® 293 Serum-free Feed Medium HA02	HA000-N011	100 mL Liquid
	HA000-N012	1000 mL Liquid
	HA000-N021	1 L Powder
	HA000-N022	10 L Powder
	HA000-N023	50 L Powder

【Note】

1. The inoculation density is designed to achieve a transfection density of about 3.3×10^6 cells/mL, and can be adjusted based on the cell expansion rate.

2. The transfection techniques provided are informational; a Design of Experiments (DOE) approach can be utilized to establish optimal experimental design.

3. Regularly monitor viable cell density and glucose concentration to determine the optimal harvest timing based on the target protein and cell viability.

DISCLAIMER

1. Use the product according to the manual instructions. Deviations from these instructions are at the user's risk, and our company will not be responsible for any resulting product performance deviations.

2. This product is for scientific research and commercial production only and is not intended for clinical diagnosis or treatment. Users assume all risks for unauthorized use, and our company shall not be responsible for any consequences.